

## PREDICTION REPORT

# A Consensus Prediction of the Secondary Structure for the 6-Phospho- $\beta$ -D-Galactosidase Superfamily

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**ABSTRACT** Two separate unrefined models for the secondary structure of two subfamilies of the 6-phospho- $\beta$ -D-galactosidase superfamily were independently constructed by examining patterns of variation and conservation within homologous protein sequences, assigning surface, interior, parsing, and active site residues to positions in the alignment, and identifying periodicities in these. A consensus model for the secondary structure of the entire superfamily was then built. The prediction tests the limits of an unrefined prediction made using this approach in a large protein with substantial functional and sequence divergence within the family. The protein belongs to the ( $\alpha$ - $\beta$  class), with the core  $\beta$  strands aligned parallel. The supersecondary structural elements that are readily identified in this model is a parallel  $\beta$  sheet built by strands C, D, and E, with helices 2 and 3 connecting strands (C + D) and (D + E), respectively, and an analogous  $\beta$ - $\alpha$  unit (strand G and helix 7) toward the end of the sequence. The resemblance of the supersecondary model to the tertiary structure formed by 8-fold  $\alpha$ - $\beta$  barrel proteins is almost certainly not coincidental. © 1995 Wiley-Liss, Inc.

**Key words:** prediction contests,  $\alpha$ - $\beta$  barrel, protein sequence alignment

A central problem in protein chemistry challenges the chemist to deduce the conformation (secondary and tertiary structure) of a protein from sequence information (primary structure). Both at the ETH in Zurich<sup>1</sup> and elsewhere,<sup>2–6</sup> progress toward solution of this problem has come through an analysis of patterns of conservation and variation in the sequences of homologous proteins.<sup>7</sup> Such an analysis is especially powerful when it is aided by detailed models of divergent evolution.<sup>8</sup> Predictions made using this approach are “consensus” models for conformation of a protein family, and assume that proteins related by common ancestry have similar conformations.<sup>9</sup>

The value of these methods can be explored by using them to make bona fide predictions, those published before an experimental structure becomes

available. To date, over a dozen bona fide predictions have been made using these methods [reviewed in refs. 7 and 10]. For about half of these, a subsequently determined crystal structure has allowed these predictions to be evaluated. In many cases, the predictions have proven to be remarkably accurate.<sup>10</sup> It is now clear that predictions are possible that miss no core secondary structural elements, misassign no  $\alpha$  helices as  $\beta$  strands (or vice versa), and do not overpredict any significant secondary structural element.<sup>11</sup> Predictions meeting these criterion are satisfactory as starting points to assemble a tertiary structural model of a protein family. Predicted secondary structures for pleckstrin homology domain,<sup>12,13</sup> hemorrhagic metalloproteinases,<sup>14</sup> and Src homology 2 domains<sup>2,3</sup> come close to meeting this standard.

Ongoing bona fide prediction efforts are necessary to define the scope of prediction methods. Over time, a large set of examples will emerge that will become statistically representative of proteins as a whole. As this set has accumulated to date, it has become clear that misassignments made by evolutionary analyses come in five principal types:

1. where multiple alignment is incorrect;
2. where the secondary structure of homologous proteins has diverged;
3. when attempting to distinguish between surface  $\beta$  strands and surface loops;
4. when attempting to distinguish between long internal  $\beta$  strands and internal helices;
5. when attempting to assign secondary structure to active site regions.

The first two (and often the first three) problems are interrelated. When the secondary structure has diverged, this often creates bad multiple alignments. Further, the distinction between a surface strand and a surface loop is often difficult even when the experimental data are in hand, and these elements

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often undergo substantial divergence in conformation during divergent evolution.

A challenge was issued on October 10, 1994, to predict the secondary and elements of the supersecondary structure of the 6-phospho- $\beta$ -D-galactosidase superfamily. The prediction was due before November 1. This protein family appeared to be an excellent target for placing the method to an extreme test. The protein is large; the target sequence has 468 amino acids. The family appears to adopt quaternary structure, at least in some cases. Both the thioglucosidase from *Brassica napus* (rape) and the thioglucosidase from *Sinapis alba* (white mustard) are reported to be homodimers, while the  $\beta$ -galactosidase from *Sulfolobus solfataricus* (not shown in the alignment) is a homotetramer. This implies that quaternary contacts might bury some residues that are on the surface of subunits, complicating the secondary structure prediction. Further, biological function has diverged substantially within the protein family, as measured by a wide divergence in substrate specificity in the member proteins. Finally, with only a few days to make a prediction, the example tests the ability of a prediction method to produce an accurate model without the benefit of extensive refinement.

A multiple alignment for the protein family was built from sequences extracted from SwissProt 29<sup>15</sup> using the DARWIN system.<sup>16,17</sup> Surface and interior residues were assigned by automated procedures similar to those described elsewhere,<sup>18</sup> the multiple alignment was parsed into units forming independent secondary structures, and elements of secondary structure were predicted within the parsed segments from patterns of conservation and variation, as described elsewhere.<sup>10,12,14,19</sup> Many of the automated routines used in this prediction are available to the public on a server accessible via electronic mail at the address [cbrg@inf.ethz.ch](mailto:cbrg@inf.ethz.ch), or using the World Wide Web (WWW) with URL <http://cbrg.inf.ethz.ch/>.

The secondary structure prediction is presented residue-by-residue in Figure 1, and summarized in Table 1. A summary of the secondary structure prediction follows:

Strand A (a009–a011; b049–051) is a short internal segment confirmed in both subfamilies.

Strand B (a014–020; b053–060) is separated from strand A by a GG dipeptide in a well anchored region of the alignment. It is largely internal. This region is interesting from a methodological point of view, as a strong assignment would not have been possible if only one of the two subfamilies were available. In both subfamilies considered alone, an internal helix would be possible. Together, however, a GG dipeptide parse at (a012–103) and a GG dipeptide parse at positions (b062–063), together with strong alignment anchoring excludes an internal helix in this region.

Parse region (a048–059; b071–081) is problematic. Subfamily b could contain a  $\beta$  strand in this region (b073–078). However, it is matched with a parsing string (PGDSC; a050–054) in sequence e of subfamily a, and a strand was not assigned.

Helix 1 (a072–084; b095–107) is reliably assigned in both subfamilies, is well anchored, and displays good amphiphilicity.

Strand C (a089–093; b111–115) is problematic in subfamily a, in part because of the small number of sequences available in this subfamily. In subfamily b, the surface and interior assignments display alternating periodicity, which confirms the strand assignment.

Active site a (a095–102; b117–125) contains conserved Arg, Ser, Trp, and Arg. It is strongly assigned.

Helix 2 (a116–130; b138–153) is reliably assigned in both subfamilies, is well anchored, and displays good amphiphilicity.

Strand D (a136–140; b159–163) is well parsed in subfamily 2, is confirmed in both subfamilies, and is largely internal.

Active site b (a141; b164–166) contains conserved Thr (part of the preceding strand) and His.

Helix 3 (a158–177; b181–198) is reliably assigned in both subfamilies, is well anchored, and displays good amphiphilicity.

Strand E (a182–185; b205–208) is assigned to a region that is near the active site, where conservation associated with active site function often obscures patterns of variation and conservation that might be used to assign secondary structure.

Active site c (a184–187; b205–209) is relatively weak, and is based ultimately on a single conserved Asn. A conserved Thr two residues before supports this conclusion. Interestingly, a Trp two residues earlier is almost completely conserved in the superfamily as well, as is a Glu immediately following the conserved Asn.

A region (b212–215) following this active site segment might be assigned as a  $\beta$  strand in subfamily b. It is not paired with a reliable assignment in subfamily a, which contains repeated parsing elements that almost certainly exclude a standard secondary structural element in this area. A similar  $\beta$  strand might be assigned in subfamily b (b219–221); this again has no corresponding element in subfamily a, and might form a  $\beta$  hairpin with the preceding strand. The alignment is poorly anchored in this region, and considerable sequence divergence between the two subfamilies is evident.

Helix 4 (a212–226; b248–268) is cleanly amphiphilic up to position a227, when an interior assignment appears on the surface arc of the amphiphilic helix. The following segment also forms a short (8 residues) amphiphilic helical pattern. In subfamily b, the helix is largely internal. Nevertheless, to the extent that amphiphilicity is detected, it

extends past the position where the amphiphilic pattern is broken. This indicates that the contacts made in subfamily a are different from those made in subfamily b. Interestingly, this helix contains a conserved His (a218; b255) and a nearly conserved His (a224; b261).

Strand x (a242–245) is cleanly parsed in subfamily a, and is canonically assigned as a short  $\beta$  strand. The segment is disrupted by parsing elements in subfamily b, which appears to be well anchored. It is possible to identify a plausible  $\beta$  segment in this subfamily. Our experience, however, has been that the experimental assignments made for such regions depend strongly on the experimental secondary structure assignment tool.

Helix x (a259–273) is not cleanly amphiphilic (position a269), but is assigned nevertheless when considering subfamily a alone. A gap is placed in its middle in subfamily b (positions b311–312). If the multiple alignment of subfamily b2 is rearranged, a helix can be detected from positions (b303–317; total length 13 positions). If the multiple alignment of subfamily b1 is adjusted, and the sequence with the deletion discarded, a weak helix can also be found. The ambiguous alignment makes all of these assignments insecure, however, and there is significant possibility that the conformations of different members of the superfamily are quite different.

Strand y (a275–280) is assigned in subfamily a only. It corresponds to a parsed region in subfamily b. Two interior residues (b323–324) might form a corresponding structure, however, in subfamily b.

The amphiphilicity of helix 5 (a286–293; b332–342) is difficult to detect when examining the alignment overall. Examining subalignments, especially of subfamily b1 and subfamily b2, makes the amphiphilicity clearer.

The region (a314) might be assigned as a short helix (7–10 residues) if the left side of subfamily a is examined alone. There is no confirmation of this helix elsewhere, however, as this region of the alignment has undergone massive sequence divergence.

Strand F (a323–327; b381–388) is badly parsed in subfamily a. The segment is conceivably a continuation of a putative helix that may follow. In subfamily b, the strand is more reliably assigned. An excellent set of anchors aligns the subalignments, and we have chosen on these grounds to make the assignment definitive in the consensus secondary structure model.

Helix y (a329–339) is short, and contains a problematic residue at position (a336). There is no confirmation for a helix assignment in subalignment b. The ambiguous alignment makes this assignment further insecure, and there is a significant possibility that the conformations of different members of the superfamily are quite different.

Strand z (a375–382) is assigned in a region of the

multiple alignment that has undergone massive sequence divergence, and where DARWIN had extreme difficulties achieving a plausible matching. It has plausible amphiphilicity in subfamily a. Therefore, the multiple alignment in subfamily b was collapsed in an effort to obtain regions that might also form  $\beta$  strands. For subfamily b1, segment (b446–452) displayed an alternating pattern. For subfamily b2, this was not possible, although it cannot be excluded that further rearrangement of the multiple alignment upon refinement could find an analogous region. As time was inadequate to do a complete search of different possible multiple alignments, no strand was assigned in this region in the consensus model.

Helix 6 (a385–398; b456–469) is well parsed, well anchored, amphiphilic, and confirmed in both subfamilies. It might, however, be missing one turn in some proteins in subfamily b.

Strand G (a404–407; b476–479) is well parsed, internal, and confirmed in both subfamilies.

Active site d (a408–410; b480–482), containing conserved Glu, Asn, and Gly, is not strongly assigned by analysis of the sequences themselves. It is, however, supported by biochemical work.<sup>20</sup>

Helix 7 (a431–448; b497–517) is well parsed, well anchored, amphiphilic, and confirmed in both subfamilies.

Residues (a451–a482; b522–554) form a remarkable segment. In subfamily b, the segment is not parsed for 35 residues, has a large number of interior residues, and apparently contains more than one secondary structural element. The first task is to parse this section. To this end, four additional columns were added to the multiple alignment by recognizing that lactase phlorizin hydrolase has multiple internal repeats. Interestingly, in two of these repeats, a parsing string PG appears. However, the repeats that contain this parsing string are cleaved proteolytically during the posttranslational modification.<sup>1</sup> These repeats are also missing Glu (b480), presumed to be part of an active site. Thus, there is no guarantee that these repeats have divergently evolved under functional constraints. This example makes an important point regarding the analysis of homologous sequences in the prediction of a protein structure.

In this region, an internal helix must be considered. Assignment of internal helices (as opposed to internal strands) relies on accurate parsing. The two subalignments were first carefully anchored. A reliable parse at (a471) was matched with a weak parse at (b541). A dipeptide GP parse in subfamily a (a460–462) was used to divide the first part of this segment. The conserved Asp was assumed to also indicate a break in secondary structure (as opposed to being an indicator of an active site position). This led to the assignment of four secondary structural elements in this region as follows:





283	YY YY	YY YY YY YY	I	parse	348	SRS —	parse	407	—	TP HH SS
284	HSS PZL	S	parse	349	TDE —	DN TT ST	408	—	DN TT ST	
285	QRD AGG	S	parse	350	RKK —	SP AA FF	409	—	SP AA FF	
286	ERK YYY	S	helix	351	GSG —	LM LM DD	410	—	LM LM DD	
287	TMT SHM	S	helix	352	SSS SKN	IT MM AA	411	—	IT MM AA	
288	LMM AHO	i	helix	353	SSS	DN DD DD	412	D	DN DD DD	
289	AEE RBR	s	helix	354	WKR ARO	A1 AA RR	413	—	A1 AA RR	
290	LGG UDF	i	helix	355	AYV AGG	NS GL G	414	—	NS GL G	
291	VVV VFF	i	helix	356	ROO ANN	—	415	SS	—	
292	RKN RRR	S	helix	357	LGL VII	V VV VV	416	SE	V VV VV	
293	EHH EBD	S	helix	358	QRK RRK	DK AA	417	MS	DK AA	
294	TII TII	i	helix	359	GGS GGS	IL SS	418	—	IL SS	
295	LII LII	i	helix	360	WV WVY	TS TT II	419	—	TS TT II	
296	USA USA	S	helix	361	GGS GGS	F1 AV	420	—	F1 AV	
297	AHE AHE	—	—	362	BOR BOR	EE ND DD	421	—	EE ND DD	
298	MNN MNN	S	parse	363	ERR ERR	HK NN RR	422	—	HK NN RR	
299	FGG KGN	S	parse	364	KSE KSE	AN SS SS	423	—	AN SS SS	
300	OSS GGN	S	parse	365	LP VEL	RR WW	424	—	RR WW	
301	DRE VII	i	parse	366	D KRN	GG PP	425	—	GG PP	
302	MUL TPT	i	parse	367	WV SPM	KI EE DS	426	—	KI EE DS	
303	P TII	i	parse	368	DL WNT	VP VS	427	—	VP VS	
304	QND AHE	S	parse	369	ZP REP	EE ND DD	428	—	EE ND DD	
305	SIL KIM	S	parse	370	ZP REP	PS GS A	429	S	PS GS A	
306	TIR ATT	S	parse	371	ZP REP	PP QP	430	L	PP QP	
307	PDD DAE	S	parse	372	Q YTH	MR VL WW	431	L	MR VL WW	
308	QES GQS	S	parse	373	IVV ILL	AA FF LL	432	Q	AA FF LL	
309	END DDD	S	parse	374	EPP QEK	SS EE MM	433	—	SS EE MM	
310	NYF DKA	S	parse	375	TRR VSS	SS EE MM	434	—	SS EE MM	
311	RKA RKA	S	parse	376	TTT SSS	TS TT TT	435	—	TS TT TT	
312	ALA ALA	—	—	377	DDD DDD	TT TT TT	436	V	TT VV	
313	ILL ILL	I	—	378	WWW WWW	IVV ILL	437	H	IVV ILL	
				379	WWW WWW	IVV ILL	438	N	IVV ILL	
				380	WWW WWW	IVV ILL	439	E	IVV ILL	
				381	WWW WWW	IVV ILL	440	E	IVV ILL	
				382	WWW WWW	IVV ILL	441	P	IVV ILL	
				383	WWW WWW	IVV ILL	442	A	IVV ILL	
				384	WWW WWW	IVV ILL	443	G	IVV ILL	
				385	WWW WWW	IVV ILL	444	—	IVV ILL	
				386	WWW WWW	IVV ILL	445	PE	IVV ILL	
				387	WWW WWW	IVV ILL	446	V	IVV ILL	
				388	WWW WWW	IVV ILL	447	T	IVV ILL	
				389	WWW WWW	IVV ILL	448	D	IVV ILL	
				390	WWW WWW	IVV ILL	449	N	IVV ILL	
				391	WWW WWW	IVV ILL	450	G	IVV ILL	
				392	WWW WWW	IVV ILL	451	W	IVV ILL	
				393	WWW WWW	IVV ILL	452	E	IVV ILL	
				394	WWW WWW	IVV ILL	453	I	IVV ILL	
				395	WWW WWW	IVV ILL	454	V	IVV ILL	
				396	WWW WWW	IVV ILL	455	P	IVV ILL	
				397	WWW WWW	IVV ILL	456	E	IVV ILL	
				398	WWW WWW	IVV ILL	457	Q	IVV ILL	
				399	WWW WWW	IVV ILL	458	G	IVV ILL	
				400	WWW WWW	IVV ILL	459	I	IVV ILL	
				401	WWW WWW	IVV ILL	460	V	IVV ILL	
				402	WWW WWW	IVV ILL	461	Y	IVV ILL	
				403	WWW WWW	IVV ILL	462	W	IVV ILL	
				404	WWW WWW	IVV ILL	463	C	IVV ILL	
				405	WWW WWW	IVV ILL	464	E	IVV ILL	
				406	WWW WWW	IVV ILL	465	I	IVV ILL	
				407	WWW WWW	IVV ILL	466	K	IVV ILL	
				408	WWW WWW	IVV ILL	467	R	IVV ILL	
				409	WWW WWW	IVV ILL	468	S	IVV ILL	
				410	WWW WWW	IVV ILL	469	F	IVV ILL	
				411	WWW WWW	IVV ILL	470	Y	IVV ILL	
				412	WWW WWW	IVV ILL	471	S	IVV ILL	
				472	WWW WWW	IVV ILL	473	G	IVV ILL	
				474	L	IVV ILL	475	P	IVV ILL	
				476	T	IVV ILL	477	I	IVV ILL	
				478	W	IVV ILL	479	M	IVV ILL	
				480	E	IVV ILL	481	N	IVV ILL	
				482	F	IVV ILL	483	G	IVV ILL	
				484	A	IVV ILL	485	A	IVV ILL	
				486	C	IVV ILL	487	C	IVV ILL	
				488	G	IVV ILL	489	G	IVV ILL	
				490	CC	IVV ILL	491	C	IVV ILL	
				492	C	IVV ILL	493	C	IVV ILL	
				494	CC	IVV ILL	495	CC	IVV ILL	
				496	CC	IVV ILL	497	CC	IVV ILL	
				498	CC	IVV ILL	499	CC	IVV ILL	
				500	CC	IVV ILL	501	CC	IVV ILL	
				502	CC	IVV ILL	503	CC	IVV ILL	
				504	CC	IVV ILL	505	CC	IVV ILL	
				506	CC	IVV ILL	507	CC	IVV ILL	
				508	CC	IVV ILL	509	CC	IVV ILL	
				510	CC	IVV ILL	511	CC	IVV ILL	
				512	CC	IVV ILL	513	CC	IVV ILL	
				514	CC	IVV ILL	515	CC	IVV ILL	
				516	CC	IVV ILL	517	CC	IVV ILL	
				518	CC	IVV ILL	519	CC	IVV ILL	
				520	CC	IVV ILL	521	CC	IVV ILL	
				522	CC	IVV ILL	523	CC	IVV ILL	
				524	CC	IVV ILL	525	CC	IVV ILL	
				526	CC	IVV ILL	527	CC	IVV ILL	
				528	CC	IVV ILL	529	CC	IVV ILL	
				530	CC	IVV ILL	531	CC	IVV ILL	
				532	CC	IVV ILL	533	CC	IVV ILL	
				534	CC	IVV ILL	535	CC	IVV ILL	
				536	CC	IVV ILL	537	CC	IVV ILL	
				538	CC	IVV ILL	539	CC	IVV ILL	
				540	CC	IVV ILL	541	CC	IVV ILL	
				542	CC	IVV ILL	543	CC	IVV ILL	
				544	CC	IVV ILL	545	CC	IVV ILL	
				546	CC	IVV ILL	547	CC	IVV ILL	
				548	CC	IVV ILL	549	CC	IVV ILL	
				550	CC	IVV ILL	551	CC	IVV ILL	
				552	CC	IVV ILL	553	CC	IVV ILL	
				554	CC	IVV ILL	555	CC	IVV ILL	
				556	CC	IVV ILL	557	CC	IVV ILL	
				558	CC	IVV ILL	559	CC	IVV ILL	
				560	CC	IVV ILL	561	CC	IVV ILL	
				562	CC	IVV ILL	563	CC	IVV ILL	
				564	CC	IVV ILL	565	CC	IVV ILL	
				566	CC	IVV ILL	567	CC	IVV ILL	
				568	CC	IVV ILL	569	CC	IVV ILL	
				570	CC	IVV ILL	571	CC	IVV ILL	
				572	CC	IVV ILL	573	CC	IVV ILL	
				574	CC	IVV ILL	575	CC	IVV ILL	
				576	CC	IVV ILL	577	CC	IVV ILL	
				578	CC	IVV ILL	579	CC	IVV ILL	
				580	CC	IVV ILL	581	CC	IVV ILL	
				582	CC	IVV ILL	583	CC	IVV ILL	
				584	CC	IVV ILL	585	CC	IVV ILL	
				586	CC	IVV ILL	587	CC	IVV ILL	
				588	CC	IVV ILL	589	CC	IVV ILL	
				590	CC	IVV ILL	591	CC	IVV ILL	
				592	CC	IVV ILL	593	CC	IVV ILL	
				594	CC	IVV ILL	595	CC	IVV ILL	
				596	CC	IVV ILL	597	CC	IVV ILL	
				598	CC	IVV ILL	599	CC	IVV ILL	
				600	CC	IVV ILL	601	CC	IVV ILL	
				602	CC	IVV ILL	603	CC	IVV ILL	
				604	CC	IVV ILL	605	CC	IVV ILL	
				606	CC	IVV ILL	607	CC	IVV ILL	
				608	CC	IVV ILL	609	CC	IVV ILL	
				610	CC	IVV ILL	611	CC	IVV ILL	
				612	CC	IVV ILL	613	CC	IVV ILL	
				614	CC	IVV ILL	615	CC	IVV ILL	
				616	CC	IVV ILL	617	CC	IVV ILL	
				618	CC	IVV ILL	619	CC	IVV ILL	
				620	CC	IVV ILL	621	CC	IVV ILL	
				622	CC	IVV ILL	623	CC	IVV ILL	
				624	CC	IVV ILL	625	CC	IVV ILL	
				626	CC	IVV ILL	627	CC	IVV ILL	
				628	CC	IVV ILL	629	CC	IVV ILL	
				630	CC	IVV ILL	631	CC	IVV ILL	
				632	CC	IVV ILL	633	CC	IVV ILL	
				634	CC	IVV ILL	635	CC	IVV ILL	
				636	CC	IVV ILL	637	CC	IVV ILL	
				638	CC	IVV ILL	639	CC	IVV ILL	
				640	CC	IVV ILL	641	CC	IVV ILL	
				642	CC	IVV ILL	643	CC	IVV ILL	
				644	CC	IVV ILL	645	CC	IVV ILL	
				646	CC	IVV ILL	647	CC	IVV ILL	
				648	CC	IVV ILL	649	CC	IVV ILL	
				650	CC	IVV ILL	651	CC	IVV ILL	
				652	CC	IVV ILL	653	CC	IVV ILL	
				654	CC	IVV ILL	655	CC	IVV ILL	
				656	CC	IVV ILL	657	CC	IVV ILL	
				658	CC	IVV ILL	659	CC	IVV ILL	
				660	CC	IVV ILL	661	CC	IVV ILL	
				662	CC	IVV ILL	663	CC	IVV ILL	
				664	CC	IVV ILL	665	CC	IVV ILL	
				666	CC	IVV ILL	667	CC	IVV ILL	

485	M	YF	XI	Y	-E	TT	OH		i
486	R	NK	NN	N	-F	PP	RR	s	parse
487	D	DD	DDD	S	-N	SS	EE	s	parse
488	E	IE	GE	G	-D	SS	ED	s	parse
489	L	VI	IV	V	-A	EE	TS	s	parse
490	V	TG	SV	E	-T	SN	DY	s	parse
491	N	FZ	LN	N	-L	RR	LL	s	parse
492	S	DN	CG	C	-P	CP	NN	s	parse
493	Q	SG	SQ	Q	-V	--	--	s	parse
494	-	KK	EE	-	-E	EQ	--	s	parse
495	-	--	--	-	-E	EQ	--	j	parse
496	-	--	--	-	-A	AA	--	s	parse
497	A	PEV	ABE	PEB	AE	DE	--	s	parse
498	I	EDD	DDD	S	EDD	RRR	--	i	helix
499	O	NSN	NNN	S	GGG	AAA	--	i	helix
500	A	EGG	EEG	T	EGG	AAA	--	i	helix
501	W	EW	WW	W	EWG	WWG	--	i	helix
502	W	WW	YY	Y	FWY	YY	--	i	helix
503	Q	KKQ	KAQ	S	KKK	QQA	--	i	helix
504	Q	GN	QA	S	KKK	QAA	--	s	helix
505	Y	YY	VV	L	YY	VV	--	i	helix
506	S	TA	TV	V	TA	TV	--	i	helix
507	S	SA	SA	S	AK	SS	--	s	helix
508	T	ST	EN	T	AK	EN	--	s	helix
509	H	NN	NN	S	NN	NN	--	s	helix
510	I	V	--	--	--	--	--	i	helix
511	I	I	--	--	--	--	--	i	helix
512	H	II	II	L	-I	II	II	s	helix
513	E	RR	RR	R	-RR	RR	--	s	helix
514	B	RR	RR	R	-RR	RR	--	s	helix
515	G	EE	DS	S	ND	YY	YY	i	helix
516	G	EE	DS	S	ND	YY	YY	s	helix
517	V	YY	VV	Y	YY	VV	--	s	helix
518	--	--	--	--	--	--	--	i	helix
519	--	--	--	--	--	--	--	s	helix
520	H	II	II	L	-I	II	II	s	helix
521	I	I	--	--	--	--	--	i	helix
522	E	RR	RR	R	-RR	RR	--	s	helix
523	B	RR	RR	R	-RR	RR	--	s	helix
524	G	EE	DS	S	ND	YY	YY	i	helix
525	G	EE	DS	S	ND	YY	YY	s	helix
526	V	YY	VV	Y	YY	VV	--	s	helix
527	--	--	--	--	--	--	--	i	helix
528	--	--	--	--	--	--	--	s	helix
529	H	II	II	L	-I	II	II	s	helix
530	I	I	--	--	--	--	--	i	helix
531	E	RR	RR	R	-RR	RR	--	s	helix
532	B	RR	RR	R	-RR	RR	--	s	helix
533	G	EE	DS	S	ND	YY	YY	i	helix
534	G	EE	DS	S	ND	YY	YY	s	helix
535	V	YY	VV	Y	YY	VV	--	s	helix
536	--	--	--	--	--	--	--	i	helix
537	--	--	--	--	--	--	--	s	helix
538	H	II	II	L	-I	II	II	s	helix
539	I	I	--	--	--	--	--	i	helix
540	E	RR	RR	R	-RR	RR	--	s	helix
541	B	RR	RR	R	-RR	RR	--	s	helix
542	G	EE	DS	S	ND	YY	YY	i	helix
543	G	EE	DS	S	ND	YY	YY	s	helix
544	V	YY	VV	Y	YY	VV	--	s	helix
545	--	--	--	--	--	--	--	i	helix
546	--	--	--	--	--	--	--	s	helix
547	H	II	II	L	-I	II	II	s	helix
548	I	I	--	--	--	--	--	i	helix
549	E	RR	RR	R	-RR	RR	--	s	helix
550	B	RR	RR	R	-RR	RR	--	s	helix
551	G	EE	DS	S	ND	YY	YY	i	helix
552	G	EE	DS	S	ND	YY	YY	s	helix
553	V	YY	VV	Y	YY	VV	--	s	helix
554	--	--	--	--	--	--	--	i	helix
555	--	--	--	--	--	--	--	s	helix
556	H	II	II	L	-I	II	II	s	helix
557	I	I	--	--	--	--	--	i	helix
558	E	RR	RR	R	-RR	RR	--	s	helix
559	B	RR	RR	R	-RR	RR	--	s	helix
560	G	EE	DS	S	ND	YY	YY	i	helix
561	G	EE	DS	S	ND	YY	YY	s	helix
562	V	YY	VV	Y	YY	VV	--	s	helix
563	--	--	--	--	--	--	--	i	helix
564	--	--	--	--	--	--	--	s	helix
565	H	II	II	L	-I	II	II	s	helix
566	I	I	--	--	--	--	--	i	helix
567	E	YY	YY	K	--	--	--	s	helix
568	W	WW	WW	W	--	--	--	s	helix
569	F	YY	YY	Y	--	--	--	s	helix
570	K	QR	RS	S	--	--	--	s	helix
571	Q	GN	GA	S	--	--	--	s	helix
572	M	YY	VV	L	--	--	--	i	helix
573	M	II	IV	V	--	--	--	i	helix
574	A	KK	SS	S	--	--	--	s	helix
575	P	EN	RN	-	--	--	--	s	helix
576	N	NN	NN	-	--	--	--	s	helix
577	--	--	--	--	--	--	--	i	helix
578	--	--	--	--	--	--	--	s	helix
579	--	--	--	--	--	--	--	s	helix
580	H	II	II	L	-I	II	II	s	helix
581	I	I	--	--	--	--	--	i	helix
582	E	YY	YY	K	--	--	--	s	helix
583	W	WW	WW	W	--	--	--	s	helix
584	F	YY	YY	Y	--	--	--	s	helix
585	--	--	--	--	--	--	--	i	helix
586	--	--	--	--	--	--	--	s	helix
587	H	II	II	L	-I	II	II	s	helix
588	I	I	--	--	--	--	--	i	helix
589	E	YY	YY	K	--	--	--	s	helix
590	W	WW	WW	W	--	--	--	s	helix
591	F	YY	YY	Y	--	--	--	s	helix
592	--	--	--	--	--	--	--	i	helix
593	--	--	--	--	--	--	--	s	helix
594	H	II	II	L	-I	II	II	s	helix
595	I	I	--	--	--	--	--	i	helix
596	E	YY	YY	K	--	--	--	s	helix
597	W	WW	WW	W	--	--	--	s	helix
598	F	YY	YY	Y	--	--	--	s	helix
599	--	--	--	--	--	--	--	i	helix
600	--	--	--	--	--	--	--	s	helix
601	H	II	II	L	-I	II	II	s	helix
602	I	I	--	--	--	--	--	i	helix
603	E	YY	YY	K	--	--	--	s	helix
604	W	WW	WW	W	--	--	--	s	helix
605	F	YY	YY	Y	--	--	--	s	helix
606	--	--	--	--	--	--	--	i	helix
607	--	--	--	--	--	--	--	s	helix
608	H	II	II	L	-I	II	II	s	helix
609	I	I	--	--	--	--	--	i	helix
610	E	YY	YY	K	--	--	--	s	helix
611	W	WW	WW	W	--	--	--	s	helix
612	F	YY	YY	Y	--	--	--	s	helix
613	--	--	--	--	--	--	--	i	helix
614	--	--	--	--	--	--	--	s	helix
615	H	II	II	L	-I	II	II	s	helix
616	I	I	--	--	--	--	--	i	helix
617	E	YY	YY	K	--	--	--	s	helix
618	W	WW	WW	W	--	--	--	s	helix
619	F	YY	YY	Y	--	--	--	s	helix
620	--	--	--	--	--	--	--	i	helix
621	--	--	--	--	--	--	--	s	helix
622	H	II	II	L	-I	II	II	s	helix
623	I	I	--	--	--	--	--	i	helix
624	E	YY	YY	K	--	--	--	s	helix
625	W	WW	WW	W	--	--	--	s	helix
626	F	YY	YY	Y	--	--	--	s	helix
627	--	--	--	--	--	--	--	i	helix
628	--	--	--	--	--	--	--	s	helix
629	H	II	II	L	-I	II	II	s	helix
630	I	I	--	--	--	--	--	i	helix
631	E	YY	YY	K	--	--	--	s	helix
632	W	WW	WW	W	--	--	--	s	helix
633	F	YY	YY	Y	--	--	--	s	helix
634	--	--	--	--	--	--	--	i	helix
635	--	--	--	--	--	--	--	s	helix
636	H	II	II	L	-I	II	II	s	helix
637	I	I	--	--	--	--	--	i	helix
638	E	YY	YY	K	--	--	--	s	helix
639	W	WW	WW	W	--	--	--	s	helix
640	F	YY	YY	Y	--	--	--	s	helix
641	--	--	--	--	--	--	--	i	helix
642	--	--	--	--	--	--	--	s	helix
643	H	II	II	L	-I	II	II	s	helix
644	I	I	--	--	--	--	--	i	helix
645	E	YY	YY	K	--	--	--	s	helix
646	W	WW	WW	W	--	--	--	s	helix
647	F	YY	YY	Y	--	--	--	s	helix
648	--	--	--	--	--	--	--	i	helix
649	--	--	--	--	--	--	--	s	helix
650	H	II	II	L	-I	II	II	s	helix
651	I	I	--	--	--	--	--	i	helix
652	E	YY	YY	K	--	--	--	s	helix
653	W	WW	WW	W	--	--	--	s	helix
654	F	YY	YY	Y	--	--	--	s	helix
655	--	--	--	--	--	--	--	i	helix
656	--	--	--	--	--	--	--	s	helix
657	H	II	II	L	-I	II	II	s	helix
658	I	I	--	--	--	--	--	i	helix
659	E	YY	YY	K	--	--	--	s	helix
660	W	WW	WW	W	--	--	--	s	helix
661	F	YY	YY	Y	--	--	--	s	helix
662	--	--	--	--	--	--	--	i	helix
663	--	--	--	--	--	--	--	s	helix
664	H	II	II	L	-I	II	II	s	helix
665	I	I	--	--	--	--	--	i	helix
666	E	YY	YY	K	--	--	--	s	helix
667	W	WW	WW	W	--	--	--	s	helix
668	F	YY	YY	Y	--	--	--	s	helix
669	--	--	--	--	--	--	--	i	helix
670	--	--	--	--	--	--	--	s	helix
671	H	II	II	L	-I	II	II	s	helix
672	I	I	--	--	--	--	--	i	helix
673	E	YY	YY	K	--	--	--	s	helix
674	W	WW	WW	W	--	--	--	s	helix
675	F	YY	YY	Y	--	--	--	s	helix
676	--	--	--	--	--	--	--	i	helix
677	--	--	--	--	--	--	--	s	helix
678	H	II	II	L	-I	II	II	s	helix
679	I	I	--	--	--	--	--	i	helix
680	E	YY	YY	K	--	--	--	s	helix
681	W								

**Fig. 1.** Multiple alignment of the two subfamilies of the b-phospho-strand-glucosidase, c-strand-d-phosphogalactosidase and d-phospho-sialan- $\beta$ -galactosidase. In regions where the alignment has been readjusted by hand, strand and residue assignments may not correspond to those produced by the automated computer output. These should be ignored.

Subfamily a (a b c i k o):

- a—(p11546) lacg - lacIa 6-phospho-strand-galactosidase (EC 3.2.1.85) (strand-d-phosphogalactosidase galactohydrolase). *Lactococcus lactis* (subsp. *lactis*) (*Streptococcus lacris*).
- b—(p11715) lacg - stau 6-phospho-strand-galactosidase (EC 3.2.1.85) (strand-d-phosphogalactosidase galactohydrolase). *Sapthyllocooccus aureus*.
- c—(p14695) lacg - lacca 6-phospho-strand-galactosidase (EC 3.2.1.85) (strand-d-phosphogalactosidase galactohydrolase). (p-strangal) (rbg). *Lactobacillus casei*.
- d—(p24240) ascb - ecob 6-phospho-strand-glucosidase (EC 3.2.1.86). *Escherichia coli*.
- e—(p26206) arbb - erwh 6-phospho-strand-glucosidase (EC 3.2.1.86). *Erwinia chrysanthemi*.
- f—(p11988) bgbl - ecob 6-phospho-strand-glucosidase (EC 3.2.1.86). *Escherichia coli*.

Subfamily b (d e f g h i l m n p q r):

- a—(p26208) bgla - clom strand-glucosidase a (EC 3.2.1.21) (gentiobiasse) (cellobiasse) (strand-d-glucoside glucohydrolase). *Clostridium thermocellum*.
- b—(p10482) bgls - calsa strand-glucosidase a (EC 3.2.1.21) (gentiobiasse) (cellobiasse) (strand-d-glucoside glucohydrolase) (armygdalase). *Caldocelium saccharolyticum*.
- c—(p22037) bgla - bacpo strand-glucosidase a (EC 3.2.1.21) (gentiobiasse) (cellobiasse) (strand-d-glucoside glucohydrolase) (armygdalase). *Bacillus polymyxa*.
- d—(q03505) bgla - bacci strand-glucosidase (EC 3.2.1.21) (gentiobiasse) (cellobiasse) (strand-d-glucoside glucohydrolase) (armygdalase). *Bacillus circulans*.
- e—(p22050) bgbl - bacpo strand-glucosidase b (EC 3.2.1.21) (gentiobiasse) (cellobiasse) (strand-d-glucoside glucohydrolase) (armygdalase). *Bacillus polymyxa*.
- f—(q12614) bgls - agnsp strand-glucosidase (EC 3.2.1.21) (gentiobiasse) (cellobiasse) (strand-d-glucoside glucohydrolase) (armygdalase). *Agrobacterium sp.* (strain atc 21400).
- g—(q00326) myro - brana myrosinase precursor (EC 3.2.3.1) (sinigrinase). *Brasicaceae* (rape).
- h—(p09849) iph - rabbit pos 1361 to 1926 of lactase-phlorizin hydrolase precursor (EC 3.2.1.108) (EC 3.2.1.62) (lactase-glycosylceramidase) (iph). *Oryctolagus cuniculus* (rabbit).
- i—(p29092) myr3 - sinhal myrosinase mb3 precursor (EC 3.2.3.1) (sinigrinase). *Sinapis alba* (white mustard).
- j—(p26204) bgls - trp noncyanoogenic strand-glucosidase precursor (EC 3.2.1.21). *Trifolium repens* (creeping white clover).
- k—(p09849) iph - human pos 1361 to 1927 of lactase-phlorizin hydrolase precursor (EC 3.2.1.108) (EC 3.2.1.62) (lactase-glycosylceramidase). *Homosapiens* (human).
- l—(p26205) bgbl - trp cyanogenic strand-glucosidase precursor (EC 3.2.1.21) (linamarase) (fragment). *Trifolium repens* (creeping white clover).

**Tritillium repens** (creeping white clover).  
—p26205 Bgt.—Imp. cyathophyllic stolonif. glaucous.

**TABLE 1. Consensus Secondary Structure Prediction for the 6-Phospho- $\beta$ -D-galactosidase Superfamily\***

Strand A	009-011	Strand A	049-051	
Strand B <sup>†</sup>	014-020	Strand B <sup>†</sup>	053-060	Internal
Helix 1 <sup>†</sup>	072-084	Helix 1 <sup>†</sup>	095-107	Amphiphilic
Strand C?	089-093	Strand C	111-115	Amphiphilic
Act site a	095-102	Act site a	117-125	
Helix 2 <sup>†</sup>	116-130	Helix 2 <sup>†</sup>	138-153	Amphiphilic
Strand D <sup>†</sup>	136-140	Strand D <sup>†</sup>	159-163	Internal
Act sit b <sup>†</sup>	141	Act sit b <sup>†</sup>	164-166	
Helix 3 <sup>†</sup>	158-177	Helix 3 <sup>†</sup>	181-198	Amphiphilic
Strand E	182-185	Strand E	205-208	
Act sit c	184-187	Act sit c	207-209	
Helix 4 <sup>†</sup>	212-226	Helix 4 <sup>†</sup>	248-268	Largely internal
Strand x	242-245			Ambiguous alignment
Helix x	259-273		318-320	Shifted alignment
Strand y	275-280	Helix 5 <sup>†</sup>	332-342	Amphiphilic
Helix 5 <sup>†</sup>	286-293	Strand F	381-388	Interior
Strand F	323-327	Gap		Ambiguous alignment
Helix y	329-339	Strand z	446-452 <sup>‡</sup>	Amphiphilic
Strand z	375-382	Helix 6 <sup>†</sup>	456-469	Amphiphilic
Helix 6 <sup>†</sup>	385-398	Strand G <sup>†</sup>	476-479	Internal
Strand G	404-407	Act site d <sup>†</sup>	480-482	
Act site d <sup>†</sup>	408-410	Helix 7 <sup>†</sup>	497-517	Amphiphilic
Helix 7 <sup>†</sup>	431-448	Strand H <sup>†</sup>	521-525	Amphiphilic
Strand H <sup>†</sup>	450-454	Strand I <sup>†</sup>	527-530	Interior
Strand I <sup>†</sup>	456-459	Strand J <sup>†</sup>	535-539	Interior
Strand J <sup>†</sup>	464-467	Strand K <sup>†</sup>	548-554	Interior
Strand K <sup>†</sup>	478-482	Helix 8 <sup>†</sup>	563-576	Amphiphilic
Helix 8 <sup>†</sup>	496-509			

\*Assignments in the consensus model (which applies to the entire superfamily) are designated with upper case letters A-K (for  $\beta$  strands) and Arabic numerals 1-8 (for  $\alpha$  helices). Strands and helices designated by "x," "y," and "z" are not part of the consensus model, and may be present in only some members of the superfamily. Assignments marked with "?" are weak within one subfamily, but confirm a stronger assignment in the other subfamily.

<sup>†</sup>Reliable assignments.

<sup>‡</sup>The multiple alignment is ambiguous; see text.

Strand H (a450-454; b521-525) is amphiphilic and confirmed in both subfamilies.

Strand I (a456-459; b527-530) is interior and confirmed in both subfamilies. It may be longer by two residues in subfamily b.

Strand J (a464-467; b535-539) is interior and confirmed in both subfamilies.

Strand K (a478-482; b548-554) is interior, well anchored, and confirmed in both subfamilies.

Finally, helix 8 (a496-509; b563-576) is amphiphilic, well anchored, and confirmed in both subfamilies.

In examining the consensus secondary structural model reported in Table 1, it is difficult not to notice the secondary structural pattern characteristic of an 8-fold  $\alpha$ - $\beta$  barrel protein. This tertiary structural hypothesis does not rest solely on pattern recognition. The model is, in fact, enforced by the active site assignments designated in Table 1. Here,  $\beta$  strands C, D, E, and G all must terminate near the active site of the protein, as in an 8-fold  $\alpha$ - $\beta$  barrel. While other topologies could also bring these residues together, this is our preferred tertiary structural model.

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