Volume 318, number 2, 118–124 FEBS 12125 © 1993 Federation of European Biochemical Societies 00145793/93/S6.00

с

86

March 1993

The nitrogenase MoFe protein

A secondary structure prediction

Dietlind L. Gerloff^a, Thomas F. Jenny^{a,b}, Lukas J. Knecht^b, Gaston H. Gonnet^b and Steven A. Benner^a

*Laboratory for Organic Chemistry and *Institute for Scientific Computation, ETH Zürich, CH-8092 Zürich, Switzerland

Received 14 December 1992

Surface residues, interior residues, and parsing residues, together with a secondary structure derived from these, are predicted for the MoFe nitrogenase protein in advance of a crystal structure of the protein, scheduled shortly to appear in *Nature*. By publishing this prediction, we test our method for predicting the conformation of proteins from patterns in the divergent evolution of homologous protein sequences in a way that places the method 'at risk'.

Protein structure prediction; Secondary structure prediction; Nitrogenase

1. INTRODUCTION

We have recently developed procedures for extracting conformational information from patterns in the divergence and conservation in the sequences of homologous proteins [1]. These procedures are based on models for the divergent evolution of behavior and structure of proteins [2–4]. The procedures have been used to predict various aspects of the conformation of several protein families [1,5]. In the cases of protein kinase [6] and the Src homology domain 3 [7,8], secondary structure predictions were made before crystallographic data became available and shown to be remarkably accurate by subsequently determined crystal and NMR structures [9– 12].

The best way to test the power of structure prediction procedures is to apply them to make predictions in advance of experimental information concerning conformation. To be useful, the predictions must be published. This ensures that knowledge of the structure cannot bias the prediction, the predictions (both correct and incorrect) are visible, and the method is placed 'at risk'. The only problem is one of coordination. A prediction published years in advance of an experimental structure is uninteresting. A prediction made even days after a structure becomes available to the predictor is useless.

In the October 29, 1992 issue of Nature [7], we invited scientists to send sequences to use as prediction targets for our procedure for proteins (a) the structure of which shortly will be solved, (b) where no structure is available

for any obviously homologous protein, (c) where a set of homologous sequences are available, (d) where these sequences are sent to us by computer mail together with a few literature citations that provide an overview of the chemistry and biology of the protein family, and (e) when enough time is available to allow coordination of the publication of the prediction and publication of the structure. This Letter reports our first efforts directed towards this end.

Our first task has been to address challenges where criterion (e) was not fully met. For example, on November 16, Prof. D.C. Rees from the California Institute of Technology challenged us to predict a secondary structure for the MoFe protein of nitrogenase. He noted that the crystal structure of this protein had been solved, and that a manuscript coauthored with J. Kim describing that structure was in press in *Nature*, scheduled to appear in the week of December 14, 1992.

Four weeks is insufficient time to assemble a complete model for the conformation of any protein family. Nevertheless, the nitrogenase is an extremely interesting target. It is a large protein and it plays a critical role in an important metabolic process. Therefore, we have used the available time to assemble a first stage prediction of the secondary structure of this protein family. The prediction turns out to be especially instructive for those seeking to apply our procedures to their own proteins. Further, when this Letter appears in print, the issue of *Nature* containing the crystal structure will be in the library, and the success of the prediction can be immediately determined.

2. RESULTS

In presenting this prediction, we address one criticism

Correspondence address: S.A. Benner, Laboratory for Organic Chemistry, ETH Zürich, CH-8092 Zürich, Switzerland. Fax: (41) (1) 262 2437.

FEBS LETTERS

of our procedure transmitted to us by established workers in the area: that it is inferior because it is not fully automated, and relies in part on the experimence and training of individuals making the prediction. As noted elsewhere, we do not find this criticism particularly evincing [1,2,6,12]. Conformational analysis in proteins is not fundamentally different from conformational analysis in other branches of organic chemistry, and no predictive problem in conformational analysis in chemistry has yet been solved, even for small molecules, by a fully automated procedure in the century during which conformational analysis are solved in chemistry by first developing a formalism. The formalism is then applied by humans to real problems. In this application, experience, training and intuition can make contributions, errors can be understood, and the formalism can be rationally improved. Organic chemical analyses can be taught, reproducibly applied, any subjected to critical testing, as any student in an undergraduate chemistry course can confirm. Of course, it is difficult to apply methods designed to evaluate automated prediction heuristics to the prediction heuristics obtained by an organic chemical paradigm. This is one reason why de novo predictions, such as the one presented here, are so important in developing the predictive formalism.

To illustrate this point, the prediction in Fig. 1 is broken into several parts. For surface, interior, parsing, and active site assignments, the first line (TJ) reports



March 1993

FEBS LETTERS

March 1993

														5110									Ivia
			20		1	215	220	22	5 23	0 23	5 240	245		255		265	270	275	280	285	290	295	300
	a			VAVR			Å		EA .	•		DKINL	LTG	WVNP	I CDVKFI		l SEMDIE		1	ED CD	ILPD GS		
	P			ECVK			TE	AHGKO	30		PS	GKLNV	FPG	_wv <u>np</u>	GDAART	KRYFI	KEMDVE	ANIY	HDTED	FDSPI	MLPN_KS	SIFTH	GRT
	d			NMMK			Т	EG	к	KAT	SN	GKINF	IPGED	ΤΥ να	CNNRFT	KRMMC	WACUD	VTT1 0	-Deco	reneral			0.00
	с х)			NMIK NMMK			w	DOVA		ENFDT													
		- \	TGYD	NALK	GIL	EHF		DGK	AGTA	ALERKI PKLERM		CUTHL	10010	61 IV <u>(</u>	JUNKEI	NKLES	LMNVII	YT11.0	DGSDV	WDTP	A <u>DG</u> E_YA A <u>DG</u> E_FR I <u>DG</u> E_FR	DWAU 1	CCT
	f	- 1	TGYD	NALK	GIL	EHF	W		AGTA	PKLERN	PN												
				NMFE			T	DYOGO	LI DOK	KSM <u>DD</u> H PK1	VVGSN		ATOLE	1112 9	<u> 201</u> F R V I	NKMLD	567676	ISLLS	SDPEEV	/LDTPJ	ADCO FR	2MYA 4	CCT
	i q	- v	TGWD	NMFE	GFA	KTF	?A	DYQGQ	PGKL			_KLNL	VIGEE	TYL C	MFRVL	KRMME	QMAVP	CSLLS	DPSEV	LDTP!	ADGQ_YR ADGH_YR	NYS 0	GGT
	-							NTGAK				GKINV		V <u>GP</u> A	DMREI	KRLFE	AMDIP	Y IMF <u>P</u>	DTSG	LDGP	TGE_YK	MYPE	<u>IG</u> T
	m	- E	EGWA	KAVT	MIN	A I TF	IGE	A <u>NG</u> LR QE R	DP			<u>S</u> KVNV RTIAI	LPGCH	_LTPG	DLDEL	RALLE	DFGLY	<u>es</u> fl <u>p</u>	DLAGS	LDGHI	PDE_FT	STTI	GG I
		~ Q	DGWA	KAVE/	\IV/	ALVF	-VTA	e Tr	DPDL			ROVTL			EIDEA	VRMIR	AFGLS	PIILP	DLSGS	LDCU	PDDRWV	PTTY	I DC
		- E	SGFA		AIVE	TLVP	ERRD	QVGKR RPAQR	P			RQVNV		LTPG	DLEYI	AESIE	SFGLRI	PLLIP	DLSGS	LOCHT	DENDEN	AT TTO	201
	1.						P	RPAQR				RRVNL	LVSHL	_CSPG	DIEMT	RRCVE	AFGLQ	PIILP	DLAQS	MDGHL	AOGDES	PLTO	GT
	2.TJ 3.SB	1	spiss	siisi SIISS	iis	sipp	ppsp	opppp	ррррг	ppppp	ppppp	o Sisii	ppppp	ppppi	sissis	sslis:	sisisi	is111	sissp	lippi	iispis	Lison	op i
	4.SB	!	1																		ssSPIS.		1
	5.DG 6.TJ	A A	SP154		IIS	IPP	PPPPI	pppp	PPPPP	PPPPP	PPPPPP	SISI	PPPPP	РРРРр	ssSSIs	5511S	SISIS	11 .	s ssP	1\$pps	ppp spSPsS:	i sPp	ip ip/
	7.TJ	b													ΛΛΑΛ <i>Ι</i>	AAAA	AAA AA BBBBBBB	AAAA	AAAA				
	8.DG 9.DG			AAAA	AAA	AA									ΑΑΑΑ	AAAA			ubb				
	10.5B			AAAA	AAA	AA						BBBBB	•				BBBB b	bb					
	11.SB	2										BBBBB			лалая	AAAA/ F	4AA 388888	bbb					
	12.00	h Ał	AAAA	AAAA	AAA	AA	unass	igneo	d due	to gi	ŧр	BBBBB	cccc	ccccc	λλαλά	\AAAA/	AAA		C	cccc	ccccc	ccc	с
			305		0 3	315	320	325	330	335	340	345	350	355	360	365	370 :	375	380	385	390 3	95 4	100
	a -	TI	EDLI	ן DT <u>GN</u> /	ARAT	IFALI	I IRYEG	TKAAE	YLOKI	FEIPA		10 TO L	1 C 1 D M T		1		POSLA	.•	1		1	•	1
	р-	ΤV	EDIA	DSAN	ALA1	rlsl <i>i</i>	RYEG	NTTGE	LLOKT	FAVPN	ALV	NTPY	GIKNT	DMLRK	IAEVT	G_KPI G_KEI	PESLV	4 2	ERGVA	IDALA LDALA	DLTHMF	LAEKE	RV evi
	o -									FAVPN		NTPY	SIKNT	DMLRK	IAEIT	G_KEI	PESLV				DLAHME		
	р с-	NL PL	EDAA	DSINA	AKAT	TENC	AYTT	PKTRE	YIKTO	WKQET	ov	LRPF	SVKGTL	DEFLTA	VSELT	G_KAI	PEELEI		ERGRL	VDAIT	D SYAW	THGKK	(F
	×x-	TF	AEAE,	AALNA	каз	IVCMC	GIST	EKTMA	YIOER	GOEVV	AL.	NTPMU HCP10	SVIGIL SVIGIL	DELLMK	VSCIS	G_KPS	RGVKL SEELK		ERGRL	VDAIA	D_SHTH	LHCKF	₹F
	e -	TL	KDAAI	AHIAN	кал	ISMC	OWCT	EKTLS	FAAEH	IGODVL	SF	NYPVO	LSAT	DFIVA	LSRIS	G KEI	PEQLAR		ERGRL	VDAIG	T_SISY D_SSAH	VHCKN	(F
	f - d -	TO	EDTAI	VAIHA	AKAT	VILO	QWCT	EKTLP	FVSEH	GODVV	SF	_NYPVC	JVSAT	DLLVA	LSRISC	G KEII	PEQLAR	2	ERGRL	VDAIA	D SSAH	IHCKK	(F
	h -	ΤQ	QEMKI	EA <u>PD</u> A	IDT	LLLC	PWQLI	KSKK	VVOEM	WNOPA	rev	AIPLO	SLAATC	DELLMK	VSOLSO	<u>с_орн</u> с_кри	PASLTH ADALTI		ERGRL	VDMMT	D <u>s</u> htwi D Shtwi	LHGKR	IF
	1 -									WNQPA		AIPLO	LAATE	ELLMT	VSOLS	G_KPI/	ADALTL				D_SHTWI		
	g k									CKVPF							PASIEE				D_AQQYI		
	- m -	SVI	EEIRE	LGTA	AOC	IAIG	A_QM(E_HMF	RAAE	VMQTK FMKTL	TGVPF	۹۷F	ERLCO	LHPNC	DEMME	LSEISC	_RPI	PSKYRR		ORSOL	ADAML	D_AHEH1	I <u>GG</u> RK	v
	n -	RL)DI AF	IPRS	AVT	LAIG	E QMF	(AAAP!	4IEDR	ALVPY	VF	QSLTC	LKVVD	AFVRV	LSSISC	I_RPAE	PAKVRR P <u>PS</u> TKR		RRAQLO	DALL	D_GHFHS D_AHFF1	SAGKK	I
	1-	57.	FLA	AGQS	VAT	LVVG	O SLA	GAAD	ALAER	TGV <u>PD</u> CRGEV	RF	GMLYC	LDAVD	AWLMAI	LAEISC	NPVE	DRYKR		DRAQL	DAML	D THEMI	LSSAR	т
	1.	•••										PHLMI	LERCD	AP1HQ1	LAKI <u>SC</u>	<u>_</u> RAVI	PEWLER				D_СНММГ		
	2.TJ 3.SB	Sis	5 S S S S	1151	s1 i	isii	spsis	ssis	11555	1	000	nnitt	4 - 4 4 4	- / / /							• ipiiii		
	4.58																	PPPPs	ssxli	III	SP sII1	l ssS	I
	5.DG 6.TJa	519	siss	1151	Ssi	111/	oP IS	S sSs	ISSS	SsSss	Spppp	SSpip	ppp ISs/\$	\$11551	s i IsSI/p	PsSIp	ېږ SSISS	PPPPs	\$\$\$XI	\$111	1 \$P1/111	l l//sS	1
	7.TJb			aa		aaaa bbbb	AAA	AAAA	AAAA.	A		A bbb	клала	ΑλλλΑ/	NAAAA	· ·			4444	00000	<u>د</u>		
	8.DGa	AA	AAAA	AAa			AAA	аааа,	AAAA	AAA				Алаалл	\AAAAA				2222	bbbb aaaaa	5 BB888	BBB I	в
1	9.DGb 0.SBa	a A 2	4444	AA	BI	BBBB		ААЛАА												bbb		в	в
1	1.SBb					889B							49999	λλλλλ	•^^^^				aaaa	aaaaa	a bbBBB		
)	2.con	AA	AAAA	A C	C BI	8888	AAA	алла	AAAA	A CCC	cc	сс		~~~~~		cccc	cc	с	лаал		CCBBBB		
											Fig. 1	(2nd	part)										

Fig. 1. Multiple alignment of the beta subfamily of the MoFe nitrogenase protein. Sequences are from the SwissProt protein sequence database

Fig. 1. Multiple alignment of the beta subfamily of the MoFe nitrogenase protein. Sequences are from the SwissProt protein sequence database using the DARWIN system. Underscores denote insertions and deletions. Dashes indicate sequences with insufficient similarity to permit alignment. Parsing strings (see text) are underlined. Proteins in subbranches in the evolutionary tree are denoted by blocks of sequences. Letters preceding lines indicate the nitrogenase with the following accession numbers in the database: a (PI6267); b (P00468); c (P25314); d (P07329); e (P20621); f (P00172); g (P11374); h (P09771); i (P09772); j (P10336); k (P26507); l (P08738); m (P12781); n (P19077); o (P15334); p (P16856); xx (P15052). The highest bridge in the evolutionary tree occurs at a PAM (accepted point mutation per 100 amino acid residues) distance of 173. Lines beginning with a number indicate the following. Lines 2-5: l and i designate strong and weak interior assignments. S and s designate strong and weak surface assignments. P and p designate strong and weak parsing assignments. X designates a split in polarity type. / designates a functional split. S designates a conserved functional residue potentially part of an active site string. For discussion of these terms, see ref. 6. Line 2 shows unrefined assignments made by a computer 'expert system' on an unrefined alignment omitting sequence x. Gaps arise from subsequent alignment refinement. Assignments are associated with a numerical probability (not indicated) that influenced the inferred secondary structures. Lines 3 and 4 show primary and secondary assignments were made independently and recorded on separate lines. TJa and TJb (lines 6 and 7) are α and β strand assignments. α and β assignments made by rigorous application of secondary structure assignment made by a line using the another system. One, DGb, SBa, and SBb (lines 8, 9, 10, and 11) are α and β assignments made by two experts (D.L.G. and S.A.B.) applying various heuristics by hand.

assignments made by two experts (D.L.G. and S.A.B.) applying various heuristics by hand.

Line 12: a consensus secondary structure prediction to be compared with the crystal structure when it becomes available. Symbols as above, with C designating coil/turn assignments.

FEBS LETTERS

March 199.



those made by a fully automated package that is essentially an 'expert system' attempting to reproduce assignments made by organic chemists using experience, training and intuition applying procedures described in detail elsewhere [1,2,5,6,11]. This is the first time this package has been applied. The second, third and fourth lines reflect two sets of predictions prepared independently by two experts (D.G. and S.B.). A comparison of these lines illustrates the range of assignments made when relying on the experience, training, and intuition of individual scientists.

Secondary structure predictions, derived from patterns in surface and interior assignments, are likewise assigned separately, first by a rigorously applied heuristic (TJa and TJb, for α and β assignments) and then by two experts acting independently. The final line contains the consensus of all three predictions resulting from discussion among the experts, with the computer prediction represented by an expert (T.J.) as well. Special emphasis was placed on identifying core secondary structural units, as these are the most critical in assembling a tertiary structure model. Finally, an additional sequence (labeled xx) was introduced later into the multiple alignment to illustrate the extent to which assignments might be altered by additional sequence information.

A new procedure was used to help identify 'breaks' (or 'parses') in the secondary structure of a protein. In this procedure, dipeptides in the sequence composed of Pro, Gly, Asp, Asn, Ser, or any combinations of these FEBS LETTERS

March 1993

were identified as 'parsing strings'. Further description of the use of parsing strings as indicators of breaks in secondary structure will be presented elsewhere.

3. DISCUSSION

The first stage prediction used a multiple alignment of one family (the β family) of the MoFe protein of nitrogenases only. A second stage prediction would include input from the second, more distantly homologous, α family, which aligns satisfactorily over part of the sequence. Preliminary study of the α family yielded secondary structure predictions that strongly confirm several predictions made in the first family (e.g., the α helix assigned to positions 131-142). The comparison does not, however, help define the conformation of the unusually structured (yet certainly important, judging by a variety of sequence features) stretch from positions 165-200.

Further, the alignment was subjected only to minimal revision. In a second stage prediction, revised versions of the multiple alignment would be considered in an effort to optimize secondary structural assignments. Further, in this first stage prediction, neither a supersecondary nor a tertiary structure was modeled, nor did we use information available regarding the active site of the enzyme, the subunit structure, or the biological function of this enzyme [13]. These procedures often help identify errors in the secondary structure prediction [6]. There was, regrettably, too little time.

A certain number of inconsistencies can undoubtedly be found in the figure, again due to a shortage of time. The authors welcome inquiries, as well as additional sequences for prediction.

NOTE ADDED IN PROOF: JANUARY 4, 1992

At the Editor's request, we have compiled recently published crystallographic data for the MoFe nitrogenase protein from *Azotobacter vinelandii* [14] in a form that allows them to be compared with a first stage prediction for the protein family (Fig. 1), completed before the crystallographic data were available. Three points are important.

First, we normally do not publish discussions of our own predictions [12] until after they have been evaluated by others. Premature evaluations by predictors of their own predictions encourage a certain type of criticism that can obscure important science, no matter how circumspect these evaluations might be. Thus, our prediction of protein kinase [6] was evaluated first by the crystallographers who solved the structure [9], by Thornton et al. [15], and then briefly by Lesk and Boswell [16]. For the SH3 domain prediction, a summary of the prediction was evaluated by Sander [17] (the prediction paper was not available to the evaluators when they made their evaluation); an editorial evaluation of the full prediction will appear simultaneously with the prediction paper [8].

Second, our central message [1] is that the organic chemist's research strategy, where a scientist actively applies a chemical formalism during the prediction process, is more likely to yield useful results than one focusing on obtaining automated computational methods. This means that methods designed to evaluated automated predictions are often deceptive when applied to predictions made using other research paradigms. With a prediction method based on a chemical formalism, it is appropriate to ask why a secondary structure assignment is correct (if it is correct), or why iti s incorrect (if it is incorrect). This is especially true for a first stage prediction (Fig. 1). Fig. 2 shows several points where the prediction was influenced by gaps, problematic alignments, ambiguous patterns in surface and interior assignments, and other issues often resolved during refinement (reference [6] discusses refinement procedures). As noted above, there was insufficient time to address any of these issues.

Third, evaluating predictions made from multiple alignments raises issues that are central to the field, not peripheral as this short note might imply. A structural model for a family of proteins does not apply exactly to any individual family member, and it is not always clear how to correlate a 'consensus' model to the conformation of an individual protein. It is clear, however, that consensus models are best evaluated using more than one experimental structure, as illustrated by the example of the SH3 domain [10,11].

Overall, the results for the MoFe nitrogenase protein are typical for a first stage unrefined prediction. Helix assignments are normally rather accurate; β -strands are less so. Problems are often encountered in unrefined predictions when assigning secondary structure near the active site (e.g. the first line of Fig. 2). Here sequence divergence is dominated by functional constraints relating to catalytic function, obscuring patterns that indicate particular types of secondary structure.

We ourselves evaluate a first stage prediction by grouping the assigned units in 7 categories: 'correct' (a predicted secondary structure unit that would not adversely affect an effort to build a tertiary structure model), 'possibly correct' (a predicted secondary structure unit whose effect on a tertiary structure model depends on context), 'wrong' (a helix assigned as a strand, tabulated as an incorrect strand assignment, or a strand assigned as a helix, an incorrect helix assignment), 'missed significant' (a helix or strand not identified in a region that does not contain a gap, and where the missed unit is important to a tertiary structural model), 'missed insignificant' (a helix or strand not identified in a region that does not contain a gap, but where the missed unit does not appear important to building a tertiary structure), 'gapped' (a helix or strand

FEBS LETTERS

March 1993

70 75 80 85 90 95 100 105 110 115 120 125 130 65 Align # TVNPAKACOPLGAVLCALGFEKTMPYVHGSQGCVAYFRSYFNRHFREPVSCVSDSMTEDAAVFGGQQ Seq ВВВВВ СВВВВВ СС ВВВ ССС АЛАЛАЛАЛАЛ ВВВВВВВ АЛАЛАЛАЛАЛАЛА CCC CCC Predict BBBBBBB AAAAAA AA ..BBB Cryst | 100 • ا 90 • . . ł 120 110 80 70 Cryst #

140 145 150 155 160 165 170 175 180 185 190 195 200 Align # l NMKDGLQNCKATY_KPDMIAVSTTCMAEVIGDDLNAFINNSKKEGFI_ PDEFPVPFAHTPSFVGSH ccccc CC CCA Seq CC Predict AAAAAAAAAA CC BBBBBBBBB A Cryst • ٠ . 190 170 180 160 140 150 Cryst #130

205 210 215 220 225 230 235 240 245 250 255 260 265 270 I I I I I T____LKSMDDKVVGSNKKINIVPGFETYL_GNFRVIKRMLSEMG Align # 1 VTGWDNMFEGIARYF_ ВВВВВ СССССССС ААААААААААА Seq Predict AAAAAAAAAAAAAAA unassigned due to gaps А АААААААААААААА BBBBBBB аалаалаалаааааа A Cryst . 240 220 230 210 200 Cryst #

275 280 285 290 295 300 305 310 315 320 325 330 335 340 Align # VGYSLLSDPEEVLDTPADGQ_FRMYA_GGTTQEEMKDAPNALNTVLLQPWHLEKTKKFVEGTWKHEVPKL Sea EVLDTPADGO FRMYA GUTUELEMADARMANA VALGE МАЛААЛАЛАЛАЛА ССССС СССССССССССС СССС АЛАЛАЛА СС ВВВВВ АЛААЛАЛАЛАЛА ВВВВВВ АЛААЛАЛАЛА ВВВВВ АЛААЛАЛАЛА ВВВВВВ Predict АЛААААААА ВВВВВ BBBBBBB AAAA Cryst . 1 280 300 310 290 260 270 . Cryst # 250

345 350 355 360 365 370 375 380 385 390 395 400 405 410 Align # ERGRLVDMMTD_SHTWLHGKRFALWGDPDFVM C AAAAAAAAACCBBBBB 9BBB CCC NIPMGLDWTDEFLMKVSEISG_QPIPASLTK_ CC AAAAAAAAAAA CCCCCC Seq Predict ААААААААААА ААААА ВЕЗВВВВ ААААА сc AAAAA алалалалалалала Cryst В 370 350 360 340 330 320 Cryst #

415 420 425 430 435 440 445 450 455 460 465 470 475 480 Align # 415 420 425 436 436 4 I I I GLVKFLLELGCEPVHILCH, NGNKRWKKAVDAI AAAAAAAAA BBBBB CCCC unassigned AAAAAAAA BBBBBBB AAAAAAAAA • YGKNATVYIGKDLWHLRSLVFTD _LAASP_ АААААААА С Seq *gaps* BBBBBB tο due Predic AAAAAAAAA AAA ААААААААА ВВВВВВВВ Cryst 1 420 430 410 Cryst # 380 400 390

Fig. 2. The sequence of the MoFe nitrogenase protein from Azotobacter vinelandii, numbered according to the multiple alignment in Fig. 1, followed by the first stage, unrefined secondary structure prediction (Fig. 1) and the secondary structure assigned by crystallography [14]. A = α helix, B = β strand, C = coil or turn. Beneath is the sequence numbering of the MoFe nitrogenase protein from Azotobacter vinelandii, the protein the crystal structure of which was solved (sequence d in the multiple alignment in Fig. 1). Positions not designated A, B, or C in the prediction are left blank; non-assignments are 'canonical' in a first stage prediction whenever the multiple alignment includes a gap and whenever the 'expert' assignments disagree. See references [1], [6] and [12] for further discussion of canonical assignments in a first stage prediction and procedures used for refining these predictions.

FEBS LETTERS

March 1993

 Table I

 Secondary structure of the MoFe nitrogenase protein: comparison of the prediction and the crystal structure

	a helices	β strands
Correct	10	7
Possibly correct	. 0	2
Wrong	0	3
Missed significant	3	4
Missed insignificant	3	0
Gapped	2	ī
Overpredicted	0	2

not identified because of the canonical treatment of gaps [6,12]), and 'overpredicted' (a helix or strand assigned to a region left unassigned by the experimentalists). These numbers for the MoFe nitrogenase protein are collected in Table I. Note that these are preliminary assignments; precise assignments can be made only in the context of an effort to assemble a tertiary structure model, which necessarily follows refinement.

Above all, this comparison illustrates the importance of early communication between crystallographer and predictor to ensure that adequate time is available for refinement. We are unable to say how much our prediction would have been improved by refinement. However, adjustments made to the multiple alignment, a standard part of a refinement procedure, should at least have allowed detection of some of the secondary structures in the regions left unassigned due to gaps (see Fig. 2). More challenging would have been improvement of the secondary structure prediction in the region of the active site.

REFERENCES

- [1] Benner, S.A. (1989) Adv. Enzyme Regul. 28, 219-236.
- [2] Benner, S.A. and Ellington, A.D. (1990) Bioorg. Chem. Front. I, 1-70.
- [3] Gonnet, G.H. and Benner, S.A. (1991) Computational Biochemistry Research at ETH, Technical Report 154, Departement Informatik, ETH Zürich, Switzerland.
- [4] Gonnet, G.H., Cohen, M.A. and Benner, S.A. (1992) Science 256, 1443–1445.
- [5] Benner, S.A., Cohen, M.A., Gonnet, G.H., Berkowitz, D.B. and Johnsson, K., in: The RNA World (R. Gesteland and J. Atkins, Eds.), Cold Spring Harbor Press, in press.
- [6] Benner, S.A. and Gerloff, D. (1991) Adv. Enzyme Regul. 31, 121-181.
- [7] Benner, S.A., Cohen, M.A. and Gerloff, D. (1992) Nature 359, 781.
- [8] Benner, S.A., Cohen, M.A. and Gerloff, D. (1992) J. Mol. Biol., in press.
- [9] Knighton, D.R., Zheng, J., Ten Eyck, L.F., Ashford, V.A., Xuong, N.H., Taylor, S.S. and Sowadski, J.M. (1991) Science 253, 407-414.
- [10] Musacchio, A., Noble, M., Pauptit, R., Wierenga, R. and Saraste, M. (1992) Nature 359, 851-855.
- [11] Yu, H., Rosen, M.K., Shin, T.B., Seidel-Dugan, C., Brugge, J.S. and Schreiber, S.L. (1992) Science 258, 1665-1668.
- [12] Benner, S.A. (1992) Curr. Opin. Struct. Biol. 2, 402-412.
- [13] Kim, J. and Rees, D.C. (1992) Science 257, 1677-1682.
- [14] Kim, J. and Rees, D.C. (1992) Nature 360, 553-560.
- [15] Thornton, J.M., Flores, T.P., Jones, D.T. and Swindells, M.B. (1991) Nature 354, 105–106.
- [16] Lesk, A.M. and Boswell, D.R. (1992) BioEssays 14, 407-410.
- [17] Rost, B. and Sander, C. (1992) Nature 360, 540.